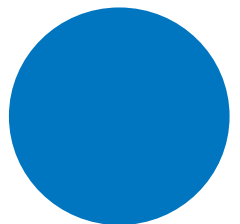
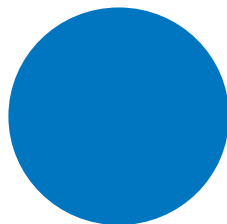
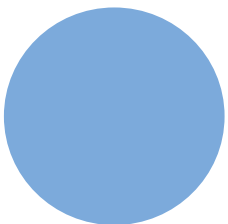
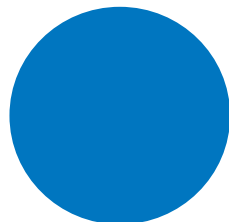
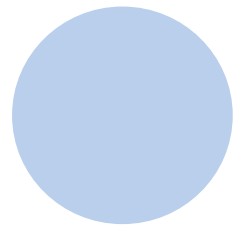
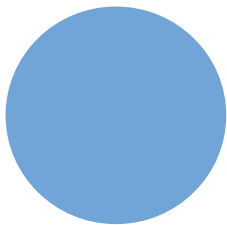
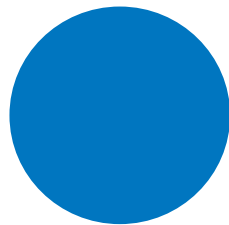
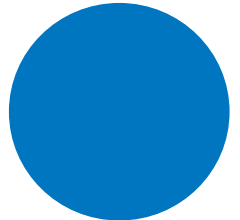
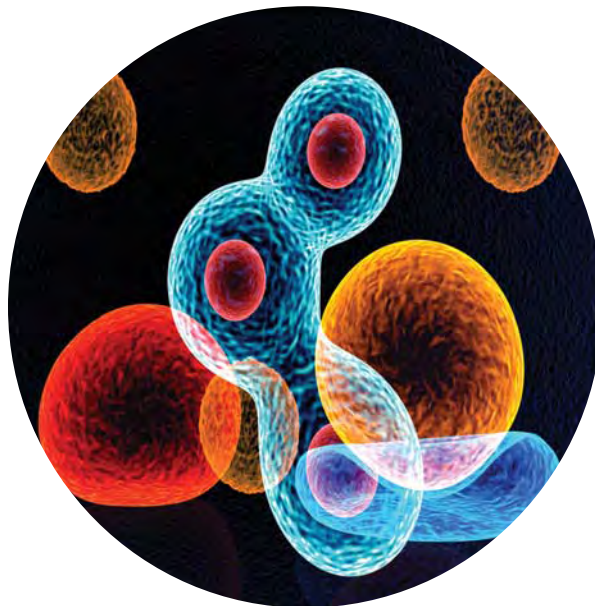
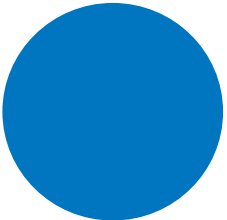
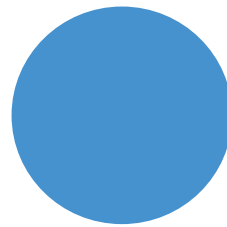
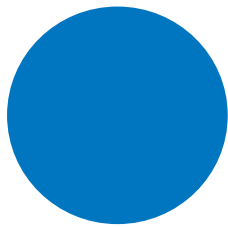
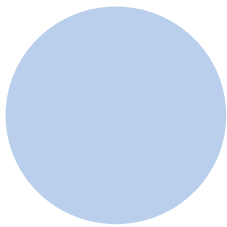
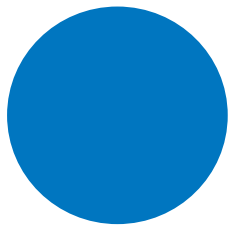


Testing Options



<p>BLOOD COLLECTION - Vacutainers Purple (Lavender) Top, EDTA</p> 	<p>STOOL COLLECTION SUPPLIES Stool Container/Lid</p> 	<p>CYTOLOGY/ MOLECULAR DIAGNOSTICS SUPPLIES Aptima Chlamydia/GC Kit, Unisex Swab</p> 	
<p>BLOOD COLLECTION – Needle Supplies Blood Culture Transfer Device- Female & Male</p> 	<p>MICROBIOLOGY SUPPLIES Blood Culture Bottle- Pediatric</p> 	<p>CYTOLOGY/ MOLECULAR DIAGNOSTICS SUPPLIES Aptima Chlamydia/GC Kit, Vaginal Swab</p> 	
<p>URINE COLLECTION SUPPLIES Sterile Plastic Cup</p> 	<p>MICROBIOLOGY SUPPLIES Blood Culture Sets-Adult</p> 	<p>CYTOLOGY/ MOLECULAR DIAGNOSTICS SUPPLIES ThinPrep Collection Vials</p> 	
<p>URINE COLLECTION SUPPLIES Urine Vacurette (IPT) Collection Tubes</p> 	<p>MICROBIOLOGY SUPPLIES e-Swab Anaerobic Transport System</p> 	<p>Collection/transport containers pictured are those provided by BBPL and are orderable online at www.bbpl.com > LabLink > Providers > Order Supplies or by calling Client Services at 800-786-4602.</p> <p>This Microbiology Testing Options list features more common tests and is not exhaustive. Visit our website Specimen Inquiry page to search our online Directory of Services for additional tests and specimen requirements.</p> <p>Collection/transport containers for most tests are not limited to those pictured. Refer to the Preferred Specimen or Other Acceptable sections of a test description within our online Directory of Services for additional acceptable containers.</p> <p>Any printed version of this document is valid only until its published date. Refer to our online Directory of Services for the most current testing information.</p> <p>Our Client Services personnel are available 24/7 to answer your questions: 800-786-4602.</p>	
<p>STOOL COLLECTION SUPPLIES Cary Blair Media</p> 	<p>MICROBIOLOGY SUPPLIES Culture Swab I (Single swab-liquid stuart media)</p> 		
<p>STOOL COLLECTION SUPPLIES O&P PVA (order O&P Set)</p> 	<p>MICROBIOLOGY SUPPLIES Culture Swab II (Dual swabs-liquid stuart media)</p> 		
<p>STOOL COLLECTION SUPPLIES O&P Formalin/ PVA Set</p> 	<p>MICROBIOLOGY SUPPLIES Culture Swab Nasopharyngeal (call to request)</p> 		
<p>STOOL COLLECTION SUPPLIES O&P Formalin (order O&P Set)</p> 	<p>MICROBIOLOGY SUPPLIES Yellow Top Vacutainer Tube, 3.32 ml, SPS</p> 		
<p>STOOL COLLECTION SUPPLIES Pinworm Paddle</p> 	<p>MICROBIOLOGY SUPPLIES M6 Red Top Viral Transport Media (HSV/RSV)</p> 		

Boyce & Bynum
Pathology Laboratories

Acid Fast Bacilli, Stain Only**Order code: 3360****Preferred specimen:** See acceptable specimen types below. Indicate source on requisition.**Sputum:**

1. A first morning, deep cough specimen is recommended.
2. If a series of 3 specimens is requested, collect specimens on 3 consecutive days at 8-24 hour intervals (24 hours when possible).
3. Collect 5.0-10.0 mL specimen (Min 3.0 mL) in tightly sealed, sterile container.
4. Refrigerate.

Urine:

1. A first morning specimen is recommended. Minimum of 10.0 mL.
2. Collect specimen in tightly sealed, sterile container.
3. Refrigerate.

Body Fluids & Bronchial Washing:

1. Submit 5.0 mL specimen (Min 1.0 mL) in tightly sealed, sterile container.
2. Refrigerate.

Spinal Fluid:

1. Submit 2.0-3.0 mL CSF (Min 1.0 mL) in a sterile screw-cap tube. Collect a separate specimen for chemistry or hematology testing if needed.
2. Submit at room temperature.

Tissue:

1. Send tissue sample in tightly sealed, sterile container.
2. Cover the tissue with sterile saline to prevent drying.
3. Keep at room temperature.

Wound Aspirates or Drainage:

1. Remove surface exudates by wiping with sterile saline or 70% alcohol. Collect fluid abscess material with a Luer tip syringe and/or remove material from the leading edge of the wound aseptically. For open lesions/abscesses, aspirate, if possible, material from under the margin of the lesion/abscess.
2. Note: Specimens submitted on swabs are not recommended. Please submit aspirate material, drainage fluid, or tissue for optimal quality of results.
3. Note: When submitting a syringe, remove the needle prior to submission and cap with a sterile syringe tip cap.
4. Refrigerate.

Feces:

1. Submit a minimum of 1 g solid stool or 1.0-5.0 mL liquid stool in a clean leak-proof container.
2. Refrigerate.

Bone Marrow & Blood:

1. Submit 5.0 mL blood (Min 1.0 mL) or 2.0 mL bone marrow (Min 0.5 mL) in either yellow (SPS) top tube or lysis-centrifugation tube.
2. Keep at room temperature.

Notes: Submit specimen in tightly sealed, sterile container.**Unacceptable:** Specimens not stored at the proper temperature, blood or bone marrow not submitted in SPS or lysis-centrifugation tubes, or specimens greater than 72 hours old.**Transport temp:** Refrigerated: Sputum, Urine, Body Fluids, Bronchial Washing, Wound Aspirates or Drainage, Feces.
Room temperature: Spinal Fluid (CSF), Tissue, Bone Marrow, Blood.**Method:** Fluorescent Stain**Unit code:** 400200**CPT Code(s):** 87206**Reported:** Within 24 hours

Use for Blood or Bone Marrow Use for Stool

Clostridium difficile Toxin by PCR**Order code: 53810****Preferred specimen:** Fresh, unpreserved, liquid or soft stool specimen in a dry, sterile screw-top container.**Minimum specimen:** 0.5 mL liquid stool or 1 g soft stool.**Other acceptable:** Stool placed in Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to Cary Blair medium using sterile swab or applicator, adding up to the fill line on the Cary Blair vial label. Mix vial well.
Stool in Para-Pak Clean or Para-Pak C&S vials is acceptable if collected and submitted according to package instructions.
Frozen raw stool is acceptable but results will be reported with a disclaimer.**Unacceptable:** Dried formed stool or stool in formalin or preservative other than Cary Blair.**Transport temp:** Refrigerated**Method:** Real-Time Polymerase Chain Reaction (PCR)**Unit code:** 538100**CPT Code(s):** 87493**Ref range:** Not Detected**Reported:** 1-2 days**Cryptosporidium Antigen-EIA****Order code: 3469****Preferred specimen:** Stool placed in 10% formalin or Cary Blair transport media.
Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to formalin or Cary Blair medium, adding up to the fill line on the transport vial label. Mix vial well. Stool specimen preserved in formalin or Cary Blair medium is stable for 7 days stored at room temperature.**Minimum specimen:** 0.5 mL liquid stool or 1 g solid (pea-sized) stool.**Other acceptable:** Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 2 hours of collection. Transport refrigerated.**Unacceptable:** Stool specimens preserved in PVA medium or multiple specimens (more than one in 24 hours).**Transport temp:** Room temperature**Method:** Enzyme Immunoassay**Unit code:** 401050**CPT Code(s):** 87328**Ref range:** Negative**Reported:** 1-3 days

Culture, Acid Fast Bacilli, with Stain**Order code: 3020****Preferred specimen:** See acceptable specimen types below. Submit specimen in tightly sealed sterile container. Indicate source on test request form.**Sputum:**

1. A first morning, deep cough specimen is recommended.
2. If a series of 3 specimens is requested, collect specimens on 3 consecutive days at 8-24 hour intervals (24 hours when possible).
3. Collect 5.0-10.0 mL specimen (Min 3.0 mL) in tightly sealed, sterile container.
4. Refrigerate.

Urine:

1. A first morning specimen is recommended. Minimum of 10.0 mL.
2. Collect specimen in tightly sealed, sterile container.
3. Refrigerate.

Body Fluids & Bronchial Washing:

1. Submit 5.0 mL specimen (Min 1.0 mL) in tightly sealed, sterile container.
2. Refrigerate.

Spinal Fluid:

1. Submit 2.0-3.0 mL CSF (Min 1.0 mL) in a sterile screw-cap tube. Collect a separate specimen for chemistry and hematology testing if needed.
2. Submit at room temperature.

Tissue:

1. Send tissue specimen in tightly sealed, sterile container.
2. Cover the tissue with sterile saline to prevent drying.
3. Keep at room temperature.

Wound Aspirates or Drainage:

1. Remove surface exudates by wiping with sterile saline or 70% alcohol. Collect fluid abscess material with a Luer tip syringe and/or remove material from the leading edge of the wound aseptically. For open lesions/abscesses, aspirate, if possible, material from under the margin of the lesion/abscess.
2. Note: Specimens submitted on swabs are not recommended. Please submit aspirate material, drainage fluid, or tissue for optimal quality of results.
3. Note: When submitting a syringe, remove the needle prior to submission and cap with a sterile syringe tip cap.
4. Refrigerate.

Feces:

1. Submit a minimum of 1 g solid stool or 1.0-5.0 mL liquid stool in a clean leak-proof container.
2. Refrigerate.

Bone Marrow & Blood:

1. Submit 5.0 mL blood (Min 1.0 mL) or 2.0 mL bone marrow (Min 0.5 mL) in either yellow (SPS) top tube or lysis-centrifugation tube.
2. Keep at room temperature.

Notes: Please indicate on test request form when the presence of *M. genavense*, *M. haemophilum*, *M. marinum*, or *M. xenopi* is suspected, as special procedures are required for isolation of these species. Culture is incubated for 8 weeks before determined to be negative. CPT code 87015 for Concentration will be added at an additional charge for body fluid, respiratory, stool, tissue, and urine specimens.

Unacceptable: Specimens not stored at the proper temperature, blood or bone marrow not submitted in SPS or lysis-centrifugation tubes, or specimens greater than 72 hours old.

Transport temp: Refrigerated: Sputum, Urine, Body Fluids, Bronchial Washing, Wound Aspirates or Drainage, Feces.
Room temperature: Spinal Fluid (CSF), Tissue, Bone Marrow, Blood.

Method: Fluorescent stain. Mycobacterial culture technique.

Unit code: 400000

CPT Code(s): 87116, 87206

Ref range: No Acid Fast Bacilli

Reported: Stain: Within 24 hours; Culture: Within 8 weeks



Use for Blood
or Bone Marrow

Use for Stool

Culture, Beta Strep, Throat**Order code: 3070**

Preferred specimen: Collect throat specimen using a sterile culture swab. Depress tongue with a sterile tongue blade and vigorously swab over each tonsillar area and the posterior pharynx while rotating the swab. Swab over any area with visual pus. Avoid touching cheeks, teeth, lips, palate and tongue.

Notes: If Group A Strep Rapid Antigen is requested, order 3069.

Unacceptable: Swab with wooden shaft, dried swab, specimen greater than 72 hours old.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 400300

CPT Code(s): 87081

Ref range: Negative for Beta Strep

Reported: Within 48 hours



or



Culture, Blood**Order code: 3080**

- Preferred specimen:** 1. Blood cultures are to be drawn into BacT/Alert culture bottles, available through BBPL Client Services or online using the BBPL Electronic Supply Order Form. All bottles should be kept at room temperature before and after collection. Always use sterile techniques with specimen collection. Sterilize the top of the blood culture bottles with alcohol and allow to air dry before injecting the blood into the bottle.
2. Adult Collection: Two bottles should be collected per draw - one aerobic with blue cap and one anaerobic with purple cap. Aerobic Media: 40 mL Tryptic Soy Agar supplemented with CO₂ and SPS. Aseptically add 5-10 mL of blood to bottle - notice gradations on side of bottle. Anaerobic Media: 40 mL Tryptic Soy Agar supplemented with SPS, reducing agents and oxygen-free nitrogen. Aseptically add 5-10 mL of blood to bottle - notice gradations on side of bottle. DO NOT OVERFILL.
3. Pediatric Collection: Pediatric bottles have a yellow cap. Each bottle contains 20 mL of BHI broth supplemented with pyridoxine. Add 1-4 mL of blood to each bottle. Only one bottle is needed for each pediatric draw.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Clotted specimens or specimens not collected in proper culture media.

Transport temp: Ambient

Method: Bacteriologic Culture Techniques/BacT/Alert 3-D

Unit code: 400500

CPT Code(s): 87040

Reported: Within 5 days



Use both for Adults

Use for Peds

Culture, Body Fluid**Order code: 3095**

- Preferred specimen:** Aspirated body fluid: Transport in a sterile leak-proof container, yellow top SPS vacutainer tube, or sterile capped syringe without needle. Submit to laboratory ASAP at room temperature.
Peritoneal dialysate fluid: If specimen will arrive in laboratory within 12 hours of collection, transport refrigerated in a sterile leak-proof container. If specimen transport time will be greater than 12 hours from time of collection, submit specimen in Blood Culture bottles. Sterilize top of bottle with alcohol and allow to air dry. Aseptically inject 10 mL fluid into a SA (blue-cap aerobic bottle) and 10 mL fluid into a SN (dark purple-cap anaerobic bottle) and transport at room temperature within 24 hours.

Notes: A Gram Stain Smear (order code 3420) must be ordered separately, if desired. Identification and susceptibility tests will be performed if indicated, at an additional charge.

Other acceptable: Large volume body fluid specimens (10 mL or more) may be submitted in blood culture bottles. Sterilize top of blood culture bottle with alcohol, allow to air dry. Aseptically inject 5-10 mL of fluid into a SA (blue-cap aerobic bottle) and 5-10 mL of fluid into a SN (dark purple-cap anaerobic bottle). Transport specimen at room temperature within 24 hours.

Unacceptable: Specimen in non-sterile container, syringe with needle, or green, purple, or blue top vacutainer tube. Specimen submitted on dried culture swab or wooden shaft swab. Frozen specimen.

Transport temp: Room temperature: Body fluid and any fluid in blood culture bottles
Refrigerated: Peritoneal fluid <12 hours old

Method: Bacteriologic Culture, Aerobic and Anaerobic.

Unit code: 400600

CPT Code(s): 87070, 87075

Ref range: No growth

Reported: Within 48 hours
Fluid submitted in blood culture bottles - within 5 days.
Broth will be held for 7 days. If the broth grows, an amended report will be issued; otherwise, no further report will be sent.



and/or

Culture, CSF**Order code: 3100**

- Preferred specimen:** Collect CSF in sterile screw-cap or snap-cap tubes. Collect 4.0-5.0 mL for adults and 0.5-1.0 mL for children. Transport to the laboratory in the collection tube **as soon as possible. Do not refrigerate.** If only one tube is collected, send to the Microbiology laboratory first. If more than one tube is obtained, the second or third collection tube should be sent to the Microbiology laboratory for culture.

Minimum specimen: 1.0 mL CSF for adults

Notes: CSF culture does not include a gram stain. If gram stain is needed order 3420 Gram Stain Smear. Identification and susceptibility tests will be performed if indicated, at an additional charge.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 401100

CPT Code(s): 87070

Reported: Negative cultures within 72 hours.
Note: Broth will be held for 7 days. If the broth grows, an amended report will be issued; otherwise, no further report will be sent.

**Culture, Ear****Order code: 3120**

- Preferred specimen:** Collect ear drainage using a sterile culture transport swab. **Do not refrigerate.** Transport swab can maintain organism viability up to 72 hours.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Dried culture swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 401200

CPT Code(s): 87070

Reported: Within 48 hours



or



Culture, Eye**Order code: 3140**

Preferred specimen: Collect eye culture using sterile culture transport swab. Corneal scrapings should be collected using a sterile corneal spatula and placed directly on appropriate culture media or placed on a sterile culture transport swab. **Do not refrigerate.** Transport swab can maintain organism viability up to 72 hours.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Dried culture swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 401400

CPT Code(s): 87070

Reported: Within 48 hours



or

**Culture, Genital****Order code: 3180**

Preferred specimen: Vaginal, cervical, urethral or penile secretions: Collect specimen using a sterile culture transport swab. Transport swab can maintain organism viability up to 72 hours. Semen or IUD: Submit in sterile sealed container. Transport ASAP within 24 hours of collection. **Do not refrigerate specimens.** Record source on test request form.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge. IUD specimens will be cultured for aerobes and for anaerobes at an additional charge.

Unacceptable: Dried culture swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 402000

CPT Code(s): 87070

Reported: IUD specimens reported after 5 days. All other specimens reported within 72 hours.



or



or

**Culture, Group B Streptococcus****Order code: 3060**

Preferred specimen: Collect both a vaginal and rectal specimen. Using a swab from a culturette container with either Amies or Stuart's media without charcoal, swab the lower vagina (vaginal introitus), followed by the rectum (ie, insert swab through the anal sphincter) using the same swab. Move swab from side to side, or rotate the swab at the collection site, allowing several seconds for absorption of organisms by the swab. Alternately, two swabs may be used, one for vaginal and one for rectal - use a double swab culturette. Place the swab/swabs back into the culturette and transport to the laboratory at room temperature as soon as possible. **NOTE** that cervical specimens are not recommended and a speculum should not be used for collection. Record source of specimen on test request form.

Notes: Patient should be checked for Group B Strep during their third trimester of pregnancy.

Other acceptable: Sterile culture swabs may be submitted in Todd Hewitt Broth available through BBPL Client Services. Broth may be incubated 35-37°C for 18-24 hours prior to transport - no longer. If broth has been incubated prior to transport, please record length of incubation on broth tube.

Unacceptable: Dried culture swabs. Swabs with wooden shafts. Specimen greater than 72 hours old.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 400400

CPT Code(s): 87081

Reported: Within 3 days



or

**Culture, Group B Streptococcus with Sensitivities****Order code: 3061**

Preferred specimen: Collect both a vaginal and rectal specimen. Using a swab from a culturette container with either Amies or Stuart's media without charcoal, swab the lower vagina (vaginal introitus), followed by the rectum (ie, insert swab through the anal sphincter) using the same swab. Move swab from side to side, or rotate the swab at the collection site, allowing several seconds for absorption of organisms by the swab. Alternately, two swabs may be used, one for vaginal and one for rectal - use a double swab culturette. Place the swab/swabs back into the culturette and transport to the laboratory at room temperature as soon as possible. **NOTE** that cervical specimens are not recommended and a speculum should not be used for collection. Record source of specimen on test request form.

Notes: Sensitivities will be performed on Group B Streptococcus isolate at an additional charge. Some patients may be allergic to penicillins and susceptibility testing of the organism is indicated. Patient should be checked for Group B Strep during their third trimester of pregnancy.

Other acceptable: Sterile culture swabs may be submitted in Todd Hewitt Broth available through BBPL Client Services. Broth may be incubated 35-37°C for 18-24 hours prior to transport - no longer. If broth has been incubated prior to transport, please record length of incubation on broth tube.

Unacceptable: Dried culture swabs. Swabs with wooden shafts. Specimen greater than 72 hours old.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 400410

CPT Code(s): 87081

Reported: Within 3 days



or



Culture, Lower Respiratory Tract**Order code: 3240**

Preferred specimen: Collect lower respiratory specimen: Sputum, tracheal aspirate, bronchial wash, bronchial alveolar lavage (BAL), or transtracheal aspirate.
For sputum, carefully instruct the patient to cough deeply (not to spit) into a sterile screw-top container. The first morning specimen is best (no 24 hour collection).
Transport at least 5 mL of sputum or fluid in a tightly sealed container.

Notes: It is strongly recommended that a gram stain be performed on all expectorated sputum specimens to determine their acceptability for culture. Please order Culture, Lower Respiratory Tract, and Gram Stain (order code 3241) unless a gram stain has already been performed by the client at the time of collection. Identification and susceptibility tests will be performed if indicated, at an additional charge.
Agents such as *Bordetella pertussis*, *Chlamydia pneumoniae*, *Corynebacterium diphtheriae*, *Legionella pneumophila*, *Mycoplasma pneumoniae*, and Acid-Fast Bacilli (*Mycobacterium tuberculosis*), and Fungus require special laboratory measures for isolation and therefore require separate orders for each specific agent.

Unacceptable: Swabs. Specimens received after 72 hours of collection. Multiple specimens (more than one in 24 hours) or frozen specimens.

Transport temp: Refrigerated

Method: Bacteriologic culture techniques

Unit code: 402600

CPT Code(s): 87070

Reported: Within 48 hours

**Culture, Lower Respiratory Tract, and Gram Stain****Order code: 3241**

Preferred specimen: Collect lower respiratory specimen: Sputum, tracheal aspirate, bronchial wash, bronchial alveolar lavage (BAL), or transtracheal aspirate.
For sputum, carefully instruct the patient to cough deeply (not to spit) into a sterile screw-top container. The first morning specimen is best (no 24 hour collection).
Transport at least 5 mL of sputum or fluid in a tightly sealed container.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.
Agents such as *Bordetella pertussis*, *Chlamydia pneumoniae*, *Corynebacterium diphtheriae*, *Legionella pneumophila*, *Mycoplasma pneumoniae*, and Acid-Fast Bacilli (*Mycobacterium tuberculosis*), and Fungus require special laboratory measures for isolation and therefore require separate orders for each specific agent.

Unacceptable: Specimens submitted on swabs. Specimens received after 72 hours of collection. Multiple specimens (more than one in 24 hours) or frozen specimens.

Transport temp: Refrigerated

Method: Bacteriologic culture techniques/Gram Stain

Unit code: 402601

CPT Code(s): 87070, 87205

Reported: Within 48 hours

**Culture, Nasopharyngeal****Order code: 3220**

Preferred specimen: Routine Nasal: Use a sterile culture transport swab to swab the anterior nares only.
Nasopharynx: Using a naso-pharyngeal swab, collect the specimen by passing through the nose into the nasopharynx. Allow swab to remain for a few seconds and carefully withdraw. Place naso-pharyngeal swab into transport tube.
Do not refrigerate. Swabs can maintain organism viability up to 72 hours.

Notes: If only interested in MRSA, order 3204 MRSA Culture.
Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Dried culture swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 402200

CPT Code(s): 87070

Reported: Within 48 hours

**Culture, Neisseria gonorrhoeae (GC) Only****Order code: 3160**

Preferred specimen: Urethral, vaginal, cervical, rectal, throat, or prostatic fluid. Collect specimen on sterile culture transport swab. **Do not refrigerate.**
Transport as soon as possible; within 24-hours, recommended.

Notes: Indicate specimen source on test request form.

Other acceptable: e-Swab Anaerobic Transport System tube at room temperature before and after inoculation.

Unacceptable: Dried culture swab, calcium alginate or wooden shaft swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 401900

CPT Code(s): 87081

Reported: Within 72 hours



Culture, Stool with Shiga Toxin 1 and 2 by EIA**Order code: 3260**

Preferred specimen: Stool placed in Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to Cary Blair medium, adding up to the fill line on the Cary Blair vial label. Mix vial well. Stool specimen preserved in Cary Blair medium is stable for 72 hours refrigerated. If multiple specimens are indicated, collect on three separate days. Record the collection date and time on each specimen vial. Cary Blair media is available through BBPL Client Services or online using the BBPL Electronic Supply Order Form.

Minimum specimen: 1.0 mL liquid stool or 1 g solid (pea-sized) formed stool.

Notes: Culture detects Salmonella, Shigella, Campylobacter, and E. Coli O157:H7 organisms. Routine stool culture includes Shiga Toxin 1 and 2 testing. Cultures for other pathogens must be ordered separately. Do not order stool culture for detection of Clostridium difficile toxin, instead order 53810 Clostridium difficile Toxin by PCR.

Other acceptable: Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 24 hours of collection. Transport refrigerated. For acceptable specimens for infants, contact the BBPL Microbiology department.

Unacceptable: Multiple specimens (more than one in 24 hours), specimens in inappropriate transport media (O&P preservatives), unpreserved stool specimens received at room temperature, frozen specimens, dried culture swabs or wooden shaft swabs.

Transport temp: Refrigerated

Method: Bacteriologic Culture Techniques and Enzyme Immunoassay

Unit code: 402710

CPT Code(s): 87045, 87046, 87427 (x2)

Reported: Within 3 days

**Culture, Throat, Routine****Order code: 3280**

Preferred specimen: Collect throat specimen using a sterile culture swab. Depress tongue with a sterile tongue blade and vigorously swab over each tonsillar area and the posterior pharynx while rotating the swab. Swab over any area with visual pus. Avoid touching cheeks, teeth, lips, palate and tongue.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Swab with wooden shaft, dried swab, specimen greater than 72 hours old.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 402800

CPT Code(s): 87070

Ref range: Normal Flora

Reported: Within 48 hours



or

**Culture, Tissue****Order code: 3340**

Preferred specimen: Send tissue sample in tightly sealed sterile container. Cover the tissue with sterile saline to prevent drying. Send to laboratory ASAP at room temperature.

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Tissue specimens in formalin.

Transport temp: Room temperature

Method: Bacteriologic Culture, Aerobic and Anaerobic

Unit code: 403300

CPT Code(s): 87070, 87075, 87176

Ref range: No growth

Reported: Within 48 hours.

Note: Broth will be held for 7 days. If the broth grows, an amended report will be issued; otherwise, no further report will be sent.

**Culture, Urine, Routine****Order code: 3300**

Preferred specimen: Collect 5 mL of urine (clean-catch, catheter, cystoscopic, or suprapubic) into a sterile screw-top container. For a clean-catch urine specimen, the patient must thoroughly clean the urethral and/or vaginal area with soap rather than disinfectant before collection. If obtaining a catheterized urine specimen, collect from the catheter line into a sterile container. Do not culture from the indwelling catheter bag. Do not culture Foley tips. Note on test request form if urine is a catheterized specimen.

Transport urine to the laboratory as soon as possible after collection. If transport will be delayed more than 2 hours, refrigerate urine specimen (stable for 24 hours refrigerated). Refrigeration is not necessary if 3-4 mL of the urine is transferred to a urine transport set (yellow top vacuette with preservative, stable for 72 hours at room temperature).

Minimum specimen: 0.2 mL urine

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Urine from a catheter bag or Foley catheter tip. Delay in transport to laboratory: greater than 24 hours for unpreserved refrigerated urine or greater than 72 hours for room temperature preserved urine. Specimens in non-sterile container or expired transport container. Frozen specimens.

Transport temp: Unpreserved urine refrigerated
Preserved urine at room temperature

Method: Bacteriologic culture techniques

Unit code: 402900

CPT Code(s): 87086

Ref range: No growth

Reported: Within 2 days



or



Preferred

Culture, Wound**Order code: 3320**

Preferred specimen: Aspirate, drainage or purulent material properly obtained from an abscess, lesion or wound and submitted in a sterile syringe without needle, or on a sterile single or double swab. Note that aspirated material is superior to a swab, and that a double swab is superior to a single swab. When anaerobes are suspected, such as with deep wounds, collection with an e-Swab Anaerobic Transport System tube is preferred. When collecting surface wound cultures, decontaminate the surrounding skin prior to collecting the specimen. Please record specific wound body site on the test request form. Stability: Specimens submitted in a sterile syringe or container are stable for 24 hours at room temperature. Swabs are stable for 72 hours at room temperature.

Minimum specimen: 0.5 mL or 0.5 gram aspirated material or swab saturated with material

Notes: Identification and susceptibility tests will be performed if indicated, at an additional charge.

Unacceptable: Dried culture swab, wooden shaft swab, frozen specimen, expired transport media.

Transport temp: Room temperature.

Note: Refrigerated specimens are not recommended for recovery of some fastidious organisms such as *Neisseria* spp.

Method: Bacteriologic culture techniques, Aerobic and Anaerobic

Unit code: 403000

CPT Code(s): 87070, 87075

Ref range: No growth

Reported: No Growth cultures are reported at 48 hours. Note: Broth will be held for 7 days. If growth appears in broth after 48 hours, an amended report will be issued; otherwise, no further report will be sent.



or



or



or

**Culture, Yeast Screen****Order code: 3740**

Preferred specimen: Collect specimen using a sterile culture transport swab. Transport to laboratory as soon as possible. Transport swab can maintain organism viability up 72 hours.

Notes: The Screen reports only presence or absence of yeast. If identification of yeast is desired, order appropriate Fungal Culture code.

Unacceptable: Dried culture swab.

Transport temp: Room temperature

Method: Bacteriologic culture techniques

Unit code: 403100

CPT Code(s): 87081

Reported: Within 48 hours



or

**Culture, Yersinia****Order code: 3760**

Preferred specimen: Stool placed in Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to Cary Blair medium, adding up to the fill line on the Cary Blair vial label. Mix vial well. Stool specimen preserved in Cary Blair medium is stable for 72 hours refrigerated.

Minimum specimen: 1.0 mL liquid stool or 1 g solid (pea-sized) formed stool.

Notes: If multiple stool specimens are indicated, collect on three separate days. Record the collection date and time on each specimen vial.

Other acceptable: Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 24 hours of collection. Transport refrigerated.

Unacceptable: Multiple specimens (more than one in 24 hours), specimens in inappropriate transport media (O&P preservatives), unpreserved stool specimens received at room temperature, frozen specimens, dried culture swabs or wooden shaft swabs.

Transport temp: Refrigerated

Method: Bacteriologic culture techniques

Unit code: 403200

CPT Code(s): 87046

Ref range: No *Yersinia* species isolated.

Reported: Within 48 hours



Enterobius Vermicularis (Pinworm) Identification**Order code: 2345**

- Preferred specimen:** Pinworm paddle. Gently press the sticky side of the paddle over the perianal surface. Specimen should be collected between 9 p.m. and midnight or immediately after arising in the morning.
- Other acceptable:** Clear cellophane scotch tape. Gently press the sticky side of the tape over the perianal surface, attach the sticky side of the tape to a glass slide, and submit in a slide holder or mailer.
- Unacceptable:** Do not use opaque or frosted (scotch) tape or slides. Do not submit on coverslips or place stool on tape or paddle. Stool specimens will not be accepted.
- Transport temp:** Room temperature
- Method:** Microscopic Exam
- Unit code:** 401920
- CPT Code(s):** 87172
- Ref range:** No Enterobius vermicularis identified.
- Reported:** 1-2 days

**Fecal Leukocyte Stain (WBC)****Order code: 2165**

- Preferred specimen:** Stool placed in PVA transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to PVA medium, adding up to the fill line on the PVA vial label. Mix vial well. Stool specimen preserved in PVA medium is stable for 7 days stored at room temperature.
- Minimum specimen:** 0.5 mL liquid stool or 1 gm solid (pea-sized) stool.
- Other acceptable:** Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 2 hours of collection. Transport refrigerated. Fresh stool specimen refrigerated is stable for 2 hours.
- Unacceptable:** Stool specimens preserved in formalin or Cary Blair medium or on swabs, fresh stool specimens stored at room temperature or frozen, multiple specimens (more than one in 24 hours).
- Transport temp:** Room temperature
- Method:** Trichrome stain/Microscopic Exam
- Unit code:** 401960
- CPT Code(s):** 89055
- Reported:** 1-3 days

**Fungal Culture****Order code: 3620**

- Preferred specimen:** **Lower Respiratory:** Collect sputum in a tightly sealed sterile 50 mL centrifuge tube or in other sterile screw-cap container. Refrigerate.
- Spinal Fluid:** Submit 2.0 mL CSF in a sterile screw-cap tube at room temperature.
- Swab:** Swab affected area with a sterile culture swab. Maintain at room temperature.
- Tissue:** Submit tissue specimen in sterile screw-cap container. Add only enough sterile saline to moisten tissue. Maintain at room temperature.
- Body Fluid, including Peritoneal and Pleural Fluids:** Submit in sterile screw-top container or in yellow (SPS) top vacutainer tube at room temperature.
- Notes:** Indicate source of specimen on test request form.
- Unacceptable:** Dried swab or leaking container.
- Transport temp:** Respiratory specimens: Refrigerated
All other specimens: Room temperature
- Method:** Fungal Culture Techniques
- Unit code:** 401500
- CPT Code(s):** 87102
- Ref range:** No yeast or filamentous fungi isolated
- Reported:** Within 4 weeks



or



or

**Fungal Culture and Stain****Order code: 3600**

- Preferred specimen:** **Lower Respiratory:** Collect specimen in a tightly sealed sterile 50 mL centrifuge tube or in other sterile screw-cap container. Refrigerate.
- Swab:** Swab affected area with a sterile culture swab. Maintain at room temperature.
- Tissue:** Submit tissue specimen in sterile screw-cap container. Add only enough sterile saline to moisten tissue. Maintain at room temperature.
- Sterile Body Fluids (other than CSF):** Submit fluid in yellow (SPS) top vacutainer tube or in a tightly sealed sterile 50 mL centrifuge tube at room temperature.
- Minimum specimen:** 1 mL fluid or 1 g solid specimen
- Notes:** Indicate source of specimen on test request form.
- Unacceptable:** Dried swab or leaking container. Frozen specimens.
- Transport temp:** Respiratory specimens: Refrigerate
All other specimens: Room temperature
- Method:** Fungal Culture Techniques/Calcofluor White Stain
- Unit code:** 401600
- CPT Code(s):** 87102, 87206
- Ref range:** Stain: No yeast or filamentous fungi seen
Culture: No yeast or filamentous fungi isolated
- Reported:** Culture: Within 4 weeks
Stain: Within 24 hours



or



or



Fungal Culture and Stain, Blood**Order code: 3602****Preferred specimen:** 7.0 mL whole blood or bone marrow in yellow (SPS) top vacutainer tube or Lysis-Centrifugation tube.**Minimum specimen:** 1.0 mL whole blood or 0.5 mL bone marrow.**Unacceptable:** Specimens in tube other than SPS or Lysis-Centrifugation. Refrigerated or frozen specimens.**Transport temp:** Room temperature**Method:** Fungal Culture Techniques/Calcofluor White Stain**Unit code:** 401620**CPT Code(s):** 87103, 87206**Ref range:** Culture negative for fungus**Reported:** Within 4 weeks
Final: Negative at 4 weeks. Positive cultures are reported as soon as detected.**Fungal Culture and Stain, CSF****Order code: 3606****Preferred specimen:** 2.0 mL CSF in sterile screw-cap tube at room temperature.**Minimum specimen:** 1.0 mL CSF**Unacceptable:** Refrigerated or frozen specimens.**Transport temp:** Room temperature**Method:** Fungal Culture Techniques/India Ink Stain**Unit code:** 401630**CPT Code(s):** 87102, 87210**Ref range:** India Ink Prep: Negative
Culture: No yeast or filamentous fungi isolated**Reported:** Stain: Within 24 hours
Culture: Within 4 weeks**Fungal Culture and Stain, Skin, Hair or Nails****Order code: 3601****Preferred specimen:** Submit skin scrapings, hair or nail specimens in sterile screw-cap container. Do **not** tape specimens to a slide or put in a moist environment (saline). There must be enough specimen to be readily visible.**Notes:** Indicate source of specimen on test request form.**Unacceptable:** Specimens in formalin or saline.**Transport temp:** Room temperature**Method:** Fungal Culture Techniques/KOH**Unit code:** 401610**CPT Code(s):** 87101, 87220**Ref range:** Stain: No yeast or filamentous fungi seen
Culture: No yeast or filamentous fungi isolated**Reported:** Within 4 weeks**Fungal Culture, Blood****Order code: 3622****Preferred specimen:** 7.0 mL whole blood or bone marrow in yellow (SPS) top vacutainer tube or Lysis-Centrifugation tube.**Minimum specimen:** 1.0 mL whole blood or 0.5 mL bone marrow.**Unacceptable:** Specimen in tube other than SPS or Lysis-Centrifugation. Refrigerated or frozen specimens.**Transport temp:** Room temperature**Method:** Fungal Culture Techniques**Unit code:** 401520**CPT Code(s):** 87103**Ref range:** Culture negative for fungus**Reported:** Within 4 weeks
Final: Negative at 4 weeks. Positive cultures are reported as soon as detected.**Fungal Culture, Skin, Hair or Nails****Order code: 3621****Preferred specimen:** Submit skin scrapings, hair or nail specimens in sterile screw-cap container. Do **not** tape specimens to a slide or put in a moist environment (saline). There must be enough specimen to be readily visible.**Notes:** Indicate source of specimen on test request form.**Unacceptable:** Specimens in formalin or saline.**Transport temp:** Room temperature**Method:** Fungal Culture Techniques**Unit code:** 401510**CPT Code(s):** 87101**Ref range:** No yeast or filamentous fungi isolated**Reported:** Within 4 weeks

Fungal Stain KOH, Genital**Order code: 2433****Preferred specimen:** Use a sterile culture transport swab to collect the genital specimen. Submit the swab at room temperature.**Other acceptable:** Specimen collected on a sterile swab and placed in 1 mL sterile saline.**Unacceptable:** Dry swab. Frozen swab.**Transport temp:** Room temperature**Method:** KOH/Microscopic Examination**Unit code:** 401933**CPT Code(s):** 87210**Ref range:** Negative for yeast**Reported:** Within 48 hours**Fungal Stain KOH, Skin, Hair, Nails****Order code: 2275****Preferred specimen:** Submit skin scrapings, hair or nail specimens in sterile screw-cap container. Do **not** tape specimens to a slide or put in a moist environment (saline). There must be enough specimen to be readily visible. Indicate source on test request form.**Unacceptable:** Specimens in formalin or saline.**Transport temp:** Room temperature**Method:** KOH/Microscopic examination**Unit code:** 401930**CPT Code(s):** 87220**Ref range:** No yeast or filamentous fungi seen**Reported:** Within 48 hours**Fungal Stain Only****Order code: 3640****Preferred specimen: Lower Respiratory:** Collect specimen in a tightly sealed sterile 50 mL centrifuge tube or in other sterile screw-cap container. Refrigerate.**Swab:** Swab affected area with a sterile culture swab. Maintain at room temperature.**Tissue:** Submit tissue specimen in sterile screw-cap container. Add only enough sterile saline to moisten tissue. Maintain at room temperature.**Sterile Body Fluids (other than CSF):** Submit fluid in yellow (SPS) top vacutainer tube or in a tightly sealed sterile 50 mL centrifuge tube at room temperature.**Notes:** Indicate source of specimen on test request form.**Transport temp:** Respiratory specimens: Refrigerate
All other specimens: Room temperature**Method:** Calcofluor White Fluorescent Stain**Unit code:** 401700**CPT Code(s):** 87206**Ref range:** No yeast or filamentous fungi seen**Reported:** Within 24 hours**Fungal Stain, CSF****Order code: 3643****Preferred specimen:** 1.0 mL CSF in a sterile screw-cap tube at room temperature.**Minimum specimen:** 0.5 mL CSF**Unacceptable:** Refrigerated or frozen specimens.**Transport temp:** Room temperature**Method:** Microscopy-India Ink**Unit code:** 401530**Ref range:** Negative India Ink Prep**Reported:** Within 24 hours

Gastrointestinal Pathogen Panel**Order code: 38070**

Preferred specimen: Stool specimen placed into Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to Cary Blair medium, adding up to the fill line on the Cary Blair vial label. Mix vial well. Cary Blair media is available through BBPL Client Services or online using the BBPL Electronic Supply Order Form.

Minimum specimen: 0.2 mL liquid stool

Notes: Test includes:
 Bacteria:
 Campylobacter
 Clostridium difficile (Toxin A/B)
 Plesiomonas shigelloides
 Salmonella
 Yersinia enterocolitica
 Vibrio
 Vibrio cholerae
 Enteroaggregative *E. Coli* (EAEC)
 Enteropathogenic *E. Coli* (EPEC)
 Enterotoxigenic *E. Coli* (ETEC) *lt/st*
 Shiga-like toxin-producing *E. Coli* (STEC) *stx1/stx2*
E. Coli O157
 Shigella/Enteroinvasive *E. Coli* (EIEC)
 Parasites:
 Cryptosporidium
 Cyclospora cayetanensis
 Entamoeba histolytica
 Giardia lamblia
 Viruses:
 Adenovirus F40/41
 Astrovirus
 Norovirus GI/GII
 Rotavirus A
 Sapovirus

Unacceptable: Specimens in inappropriate transport media or frozen specimens.

Transport temp: Refrigerated

Method: Polymerase Chain Reaction

Unit code: 538070

CPT Code(s): 87507

Ref range: Not Detected

Reported: Within 24 hours

**Giardia Antigen, EIA, Stool****Order code: 3465**

Preferred specimen: Stool placed in 10% formalin or Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to formalin or Cary Blair medium, adding up to the fill line on the transport vial label. Mix vial well. Stool specimen preserved in formalin or Cary Blair medium is stable for 7 days stored at room temperature.

Minimum specimen: 0.5 mL liquid stool or 1 g solid (pea-sized) stool.

Other acceptable: Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 2 hours of collection. Transport refrigerated.

Unacceptable: Stool specimens preserved in PVA medium or multiple specimens (more than one in 24 hours).

Transport temp: Room temperature

Method: Enzyme Immunoassay

Unit code: 402010

CPT Code(s): 87329

Ref range: Negative

Reported: 1-3 days

**Gram Stain Smear****Order code: 3420**

Preferred specimen: Two unfixed slides smeared with sample or specimen from culture swab. If accompanying a culture, two sterile swabs are recommended, one for the culture and one for the gram stain. Also acceptable, any amount of liquid specimen in a sterile screw-cap container.

Notes: Indicate source of specimen on test request form.

Unacceptable: Do not tape specimen to slide. Do not use coverslip or place another slide on top of the specimen slide.

Transport temp: Room temperature

Method: Gram Stain

Unit code: 406000

CPT Code(s): 87205

Reported: Within 24 hours

**Helicobacter pylori Antigen, Stool****Order code: 3256**

Preferred specimen: 2 g stool (thumbnail size portion) or 2.0 mL liquid stool in sterile screw-cap container. Specimen must be received in laboratory same day as collected or transport frozen.

Minimum specimen: 1 g stool or 1.0 mL liquid stool

Other acceptable: Stool specimen that has been refrigerated for less than 72 hours.

Unacceptable: Specimens in transport media. Specimens at room temperature greater than 2 hours.

Transport temp: Frozen

Method: Rapid Lateral Flow Immunoassay

Unit code: 402056

CPT Code(s): 87338

Ref range: Negative

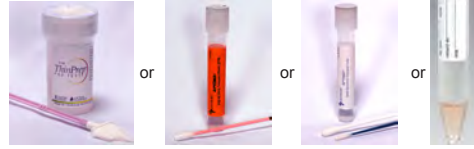
Reported: 1-2 days



Herpes Simplex Viruses (HSV) DNA

Order code: 38255

- Preferred specimen:** Collect external anogenital lesion specimen or other vesicular lesion specimen using universal transport media kit. Transport refrigerated.
Or:
 1.0 mL serum, red top tube or SST or plasma, lavender (EDTA) top tube. Remove serum or plasma from cells, transfer to a plastic transport tube and freeze. Separate specimens must be submitted when multiple tests are ordered.
- Minimum specimen:** 0.5 mL serum, plasma, CSF, SurePath media, or ThinPrep media.
- Notes:** Specimen source is required. Record source on test request form. This assay will simultaneously detect and differentiate between HSV 1 and HSV 2.
- Other acceptable:** 1.0 mL CSF in sterile container, frozen. Specimens submitted in SurePath media, ThinPrep media, or Aptima swab, refrigerated.
- Unacceptable:** Heparinized specimens, non-sterile or leaking containers.
- Transport temp:** Swab, universal transport media, ThinPrep media, or Aptima swab: Refrigerated
 All other specimens: Frozen
- Method:** Polymerase Chain Reaction
- Unit code:** 538255
- CPT Code(s):** 87530 (x2)
- Ref range:** Not Detected
- Reported:** 1-7 days



Influenza Antigen Screen A & B

Order code: 3215

- Preferred specimen:** Collect nasal swab, aspirate, wash or throat swab. Place swab in a currette container or in M6 viral transport media. Place nasal aspirate or wash in a sterile screw-cap container.
- Other acceptable:** Swab specimen in M4, M4RT, or M5 viral transport media.
- Unacceptable:** Dry swab not in currette or viral transport media.
- Transport temp:** Refrigerated
- Method:** Immunochromatographic assay
- Unit code:** 402150
- CPT Code(s):** 87400 (x2)
- Ref range:** Negative
- Reported:** Within 24 hours



Malaria, Blood Smear

Order code: 7130

- Preferred specimen:** 3.0 mL whole blood, lavender (EDTA) top tube.
- Notes:** Specimen must be received in laboratory within 24 hours of collection.
- Transport temp:** Room temperature
- Method:** Geimsa Stain
- Unit code:** 409100
- CPT Code(s):** 87207
- Ref range:** No Plasmodium/Babesia species seen.
- Reported:** 1-3 days



MRSA (ORSA) Culture

Order code: 3204

- Preferred specimen:** Specimen should be submitted in a tightly sealed sterile container or submitted using sterile culture swabs. Stool specimens: submitted in Cary-Blair transport media. Note source of specimen on test request form.
- Unacceptable:** Dried culture swab or wooden shaft swab. Specimen greater than 72 hours old.
- Transport temp:** Room temperature
- Method:** Routine Culture Technique
- Unit code:** 402330
- CPT Code(s):** 87081
- Ref range:** No MRSA isolated
- Reported:** Within 48 hours



Ova and Parasites Exam, Stool**Order code: 7140**

Preferred specimen: Stool placed in O & P transport containers, inoculate both the formalin and PVA vial. **Note:** It is very important that BOTH formalin and PVA vials be inoculated with the stool specimen. Complete analysis is only possible when both vials of preservative are submitted for testing.

Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to both the formalin and PVA vials, adding up to the fill line on each vial label. Mix vials well. Stool specimen preserved in O & P transport media is stable for 7 days stored at room temperature. If multiple stool specimens are indicated, collect on three separate days. Record the collection date and time on each specimen vial.

Minimum specimen: 0.5 mL liquid stool or 1 g solid (pea-sized) stool.

Notes: The ova and parasite exam does **not** test for the identification of *Enterobius vermicularis* (pinworm), refer to (order code 2345). Special stains are required for *Cryptosporidium*, *Cyclospora*, *Cystoisospora*, or Microsporidia and are not detected by the ova and parasite exam. For *Cryptosporidium*, *Cyclospora*, and *Cystoisospora* stains, refer to (order code 7400). Antigen testing by EIA is the preferred method for *Cryptosporidium* (order code 3469). For Microsporidia stain, refer to (order code 82692).

Other acceptable: Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 24 hours of collection. Transport refrigerated.

Unacceptable: Specimens contaminated with barium or oil, specimens preserved in Cary Blair media or on swabs, frozen specimens, or multiple specimens (more than one in 24 hours)

Transport temp: Room temperature

Method: Microscopic Examination of Stool Concentrate and Trichrome Stain

Unit code: 401910

CPT Code(s): 87177, 87209

Ref range: No parasites seen

Reported: Within 3 days

**Ova and Parasites with Giardia Antigen****Order code: 3419**

Preferred specimen: Stool placed in O & P transport containers, inoculate both the formalin and PVA vial. **Note:** It is very important that BOTH formalin and PVA vials be inoculated with the stool specimen. Complete analysis is only possible when both vials of preservative are submitted for testing.

Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to both the formalin and PVA vials, adding up to the fill line on each vial label. Mix vials well. Stool specimen preserved in O & P transport media is stable for 7 days stored at room temperature. If multiple stool specimens are indicated, collect on three separate days. Record the collection date and time on each specimen vial.

Minimum specimen: 0.5 mL liquid stool or 1 g solid (pea-sized) stool

Other acceptable: Fresh stool specimen in a clean, plastic screw-cap container is acceptable only if the specimen will be received in the testing laboratory within 2 hours of collection. Transport refrigerated.

Unacceptable: Specimens contaminated with barium or oil, specimens on swabs, frozen specimens, or multiple specimens (more than one in 24 hours).

Transport temp: Specimen in preservative: Room temperature
Fresh specimen: Refrigerated

Method: Ova and Parasites: Microscopic Examination of Stool Concentrate and Trichrome Stain
Giardia Antigen: Enzyme Immunoassay

Unit code: 401925

Ref range: Ova and Parasites: No Parasites Seen
Giardia Antigen: Negative

Reported: Reported within 72 hours

**Parasite Identification Gross Examination****Order code: 3815**

Preferred specimen: Parasite immersed in enough saline to cover specimen, submitted in a clean plastic screw-capped container.

Minimum specimen: Must be enough specimen to visually see in the container.

Other acceptable: Parasite in formalin or in a clean plastic screw-capped container.

Unacceptable: Specimens taped or mashed on a glass slide.

Transport temp: Room temperature

Method: Macroscopic and/or Microscopic Examination

Unit code: 401915

CPT Code(s): 87169

Reported: 1-3 days

**Parasitology Stain by Acid-Fast****Order code: 7400**

Preferred specimen: 5 g stool in 10% formalin vial.

Notes: *Cryptosporidium*, *Cyclospora*, and *Cystoisospora* are the only parasites detected by this stain. For other parasites, order Ova and Parasites Exam, Stool (order code 7141).

Unacceptable: Stool specimens submitted in preservatives other than 10% formalin or unpreserved stool specimens. Frozen specimens. Specimens other than stool.

Transport temp: Room temperature

Method: Modified Acid-Fast Stain

Unit code: 408000

CPT Code(s): 87015, 87207

Ref range: Negative

Reported: 1-3 days



Respiratory Pathogens Profile, Qualitative Real-Time PCR**Order code: 39000****Preferred specimen:** Nasopharyngeal swab in viral transport media.**Minimum specimen:** 1 swab in transport media or 1 mL fluid/wash.**Notes:** Profile includes: Influenza A, Influenza A Subtypes H1, H1-2009, & H3, Influenza B, Parainfluenza Types 1, 2, 3, Respiratory Syncytial Virus, Human Metapneumovirus, Rhinovirus, Adenovirus, Bordetella pertussis, Chlamydia pneumoniae, Mycoplasma pneumoniae, Coronavirus 229E, Coronavirus OC43, Coronavirus HKU1, and Coronavirus NL63.**Other acceptable:** Nasopharyngeal swab in sterile saline; nasal wash or bronchial lavage/wash in sterile container.**Unacceptable:** Dry swabs, wooden swabs, or calcium alginate swabs. Specimens greater than 72 hours.**Transport temp:** Refrigerated**Method:** Real-Time Polymerase Chain Reaction**Unit code:** 539000**CPT Code(s):** 87486, 87581, 87633, 87798**Ref range:** Negative**Reported:** 1-2 days**Respiratory Syncytial Virus (RSV) Antigen****Order code: 3470****Preferred specimen:** Nasopharyngeal swab or 0.5-1.0 mL nasopharyngeal aspirate placed immediately into M6 viral transport media or 2-4 mL nasopharyngeal wash in sterile container. Submit specimen to laboratory within 48 hours. Transport refrigerated or frozen at -20°C.**Notes:** Viral transport tube must be tightly sealed to prevent leakage. The wire from the nasopharyngeal swab cannot stick out under the screw-cap of the viral transport tube.**Other acceptable:** Nasopharyngeal swab or aspirate in M4, M4RT, M5, or normal saline (only add enough saline to cover swab).**Unacceptable:** Nasal swab in culterette tube with liquid Stuarts or Amies media.**Transport temp:** Refrigerated or frozen -20°C**Method:** Immunochromatographic Sandwich Assay**Unit code:** 402500**CPT Code(s):** 87420**Ref range:** Negative**Reported:** Within 24 hours

or

Use for Aspirate
or Wash**Rotavirus Antigen, Stool****Order code: 3460****Preferred specimen:** 1 g solid stool (pea size portion) or 0.5 mL liquid stool in a screw-cap container that is clean, dry, and free of preservatives, media and detergents. Freeze immediately after collection and submit frozen. Note that peak viral counts usually occur 3-5 days after the onset of symptoms.**Other acceptable:** Stool specimen that has been refrigerated or kept at room temperature for less than 2 hours prior to receipt in the testing laboratory.**Unacceptable:** Stool specimens containing preservatives or media, unfrozen specimens greater than 2 hours old.**Transport temp:** Frozen**Method:** Membrane Enzyme-linked Immunosorbant Assay**Unit code:** 402400**CPT Code(s):** 87425**Ref range:** Negative**Reported:** Within 24 hours

or

**Scabies (Mites) or Lice Identification****Order code: 3800****Preferred specimen:** For scabies: Dry skin scraping submitted in a dry clean plastic screw-capped container. The addition of any fluid to the container hampers recovery and identification. Do NOT tape specimen to a slide or use a coverslip. For lice or nits: Hair samples in a dry clean plastic screw-capped container.**Minimum specimen:** There must be enough specimen to visually see in the container.**Unacceptable:** Specimens submitted on slides or coverslips.**Transport temp:** Room temperature**Method:** Microscopic examination**Unit code:** 409000**CPT Code(s):** 87168**Reported:** 1-3 days**Shiga Toxin 1 and 2 by EIA, Stool****Order code: 3255****Preferred specimen:** Stool placed in Cary Blair transport media. Collect stool specimen in a clean, dry container. Immediately after collection, transfer the stool specimen to Cary Blair medium, adding up to the fill line on the transport vial label. Mix vial well. Stool specimen preserved in Cary Blair medium is stable for 14 days refrigerated. Cary Blair media is available through BBPL Client Services or online using the BBPL Electronic Supply Order Form.**Minimum specimen:** 1.0 mL liquid stool or 1 g solid (pea-sized) formed stool.**Other acceptable:** Fresh (unpreserved) stool specimen in a clean, plastic screw-cap container stored refrigerated or frozen up to 14 days.**Unacceptable:** Stool specimens preserved in PVA medium or formalin. Dry swabs or stool in diapers.**Transport temp:** Refrigerated**Method:** Enzyme Immunoassay**Unit code:** 402551**CPT Code(s):** 87427 (x2)**Ref range:** Negative for Shiga Toxin 1 and Shiga Toxin 2**Reported:** Within 24 hours

Streptococcus Group A Rapid Antigen with Reflex to Culture**Order code: 3069**

Preferred specimen: Collect throat specimen using a sterile culture swab. Depress tongue with a sterile tongue blade and vigorously swab over each tonsillar area and the posterior pharynx while rotating the swab. Swab over any area with visual pus. Avoid touching cheeks, teeth, lips, palate and tongue.

Notes: If Streptococcus Group A Rapid Antigen is negative, a culture (order code 3070) will be performed at an additional charge.

Unacceptable: Swab with wooden shaft, dried swab, specimen greater than 72 hours old.

Transport temp: Room temperature

Method: Enzyme Immunoassay/Bacteriologic Culture Techniques

Unit code: 400299

CPT Code(s): 87880

Ref range: Negative for Beta Strep

Reported: 1-2 days

**Streptococcus Group B by NAA****Order code: 3041**

Preferred specimen: Collect both a vaginal and rectal specimen. Using a swab from a culturette container with either liquid Amies or liquid Stuart's media without charcoal, swab the lower vagina (vaginal introitus) followed by the rectum (i.e., insert swab through the anal sphincter) using the same swab. Move swab from side to side, or rotate the swab at the collection site, allowing several seconds for absorption of organisms by the swab. Alternately, two swabs may be used, one for vaginal and one for rectal using a double swab culturette. Place the swab/swabs back into the culturette and transport to the laboratory refrigerated. Record source of specimen on the test request form.

Unacceptable: Cervical/endocervical, perianal, perirectal, or perineal specimens or any source other than vagina and rectum. A speculum should not be used for sample collection. Dried swabs. Frozen swabs. Swabs older than 7 days or in transport media other than Amies or Stuart's without charcoal.

Transport temp: Refrigerated

Method: Selective Broth Enrichment Culture and DNA Amplification

Unit code: 400415

CPT Code(s): 87081, 87653

Ref range: Negative

Reported: Within 48 hours

**Streptococcus Group B by NAA with Sensitivities****Order code: 3042**

Preferred specimen: Collect both a vaginal and rectal specimen. Using a swab from a culturette container with either liquid Amies or liquid Stuart's media without charcoal, swab the lower vagina (vaginal introitus) followed by the rectum (i.e., insert swab through the anal sphincter) using the same swab. Move swab from side to side, or rotate the swab at the collection site, allowing several seconds for absorption of organisms by the swab. Alternately, two swabs may be used, one for vaginal and one for rectal using a double swab culturette. Place the swab/swabs back into the culturette and transport to the laboratory refrigerated. Record source of specimen on the test request form.

Notes: If Streptococcus Group B Strep is positive, sensitivities will be added at an additional charge.

Unacceptable: Cervical/endocervical, perianal, perirectal, or perineal specimens or any source other than vagina and rectum. A speculum should not be used for sample collection. Dried swabs. Frozen swabs. Swabs older than 7 days or in transport media other than Amies or Stuart's without charcoal.

Transport temp: Refrigerated

Method: Selective Broth Enrichment Culture and DNA Amplification

Unit code: 400420

CPT Code(s): 87081, 87653

Ref range: Negative

Reported: Within 48 hours

**Vancomycin-Resistant Enterococcus Culture Only****Order code: 3203**

Preferred specimen: Stool specimen in Cary-Blair transport media. Use a sterile culture swab for collection from other sites. Record culture site on test request form.

Notes: The VRE Culture Screen reports presence or absence of VRE. If antibiotic susceptibilities are needed, order the appropriate routine culture and note "culture for VRE" on test request form.

Unacceptable: Dried culture swab or wooden shaft swab.

Transport temp: Room temperature

Method: Routine Culture Technique

Unit code: 402320

CPT Code(s): 87081

Reported: Within 72 hours

